METAL COORDINATION CORE OF BLEOMYCIN: COMPARISON OF METAL COMPLEXES BETWEEN BLEOMYCIN AND ITS BIOSYNTHETIC INTERMEDIATE

Yukio Sugiura

Faculty of Pharmaceutical Sciences, Kyoto University, Kyoto 606, Japan Received February 23, 1979

Summary: On the basis of electron spin resonance results, the 1:1 Cu(II), Co(II), Co(II)- 0_2 , and Ni(III) complexes of bleomycin(BLM) have been compared with the corresponding metal complexes of its biosynthetic intermediate(P-3A). The present study suggests that (1) P-3A is an useful ligand for the clarification of metal-binding sites of BLM; (2) the secondary amine, pyrimidine ring nitrogen, deprotonated peptide nitrogen of histidine residue, and histidine imidazole groups as planar ligand donors, and the α -amino group as axial donor, are substantially important for metal-coordination of BLM; and (3) the sugar and bithiazole portions of BLM probably contribute to stabilization of Co(II)- 0_2 adduct complex and axial sixth coordination of Cu(II) and Ni(III) complexes.

Introduction

BLM, a glycopeptide antibiotic which has been used for the treatment of selected human neoplastic disease, was originally isolated as a Cu(II) complex from fermentation broths of Streptomyces verticillus. The drug has both metal-chelating and DNA-binding sites, and its activity is presumably owing to this bifunctionality. However, the metal coordination core of BLM has never been established. The biosynthetic intermediate of BLM, P-3A, is a peptide isolated from a culture of Streptomyces verticillus and is structurally related to BLM. Herein, the coordination environment of BLM has been discussed through the comparison of metal complexes between BLM and P-3A.

Experimental

BLM-A₂ and P-3A purified were kindly supplied from Nippon Kayaku Co. Ltd. The 1:1 Cu(II) and Co(II) complexes of the antibiotics were obtained by the mixing of the ligand and metal nitrate in aqueous solution(pH 6.8), and a fully deaerated condition was kept for the preparation of the Co(II) complexes. The 1:1 Ni(III) complexes were also prepared

Abbreviations used: BLM, bleomycin; P-3A, biosynthetic intermediate of bleomycin; ESR, electron spin resonance; Li-TCNQ, 7,7,8,8-tetracyanoquinodimethane lithium salt

BLM-A2

P-3A

by the oxidation of the corresponding Ni(II) complexes with ${\rm Ir}^{\rm IV}{\rm Cl}_6^{2-.8}$ X-Band ESR spectra of magnetically dilute aqueous glasses containing the metal-antibiotic complexes(1.0 mM) were measured at 77 K using a JES-FE-3X spectrometer operating with 100 KHz magnetic field modulation. The g values were determined taking Li-TCNQ(g=2.0026) as a standard, and the magnetic fields were calculated by the splitting of Mn(II) in MgO(ΔH_{3-4} =86.9 G).

Results and Discussion

Figure 1 and Table I summarize the ESR spectral data for the Cu(II), Co(II), Co(II)-0₂, and Ni(III) complexes of the two antibiotics at pH 6.8. The average g values and A tensor of the P-3A-Cu(II) complex are close to those of the BLM-A₂-Cu(II) complex, though the spectral anisotropy(g_{zz} =2.214, g_{yy} =2.133, g_{xx} =2.078, A_{\parallel} =167 G, A_{\perp} =72 G) of the former

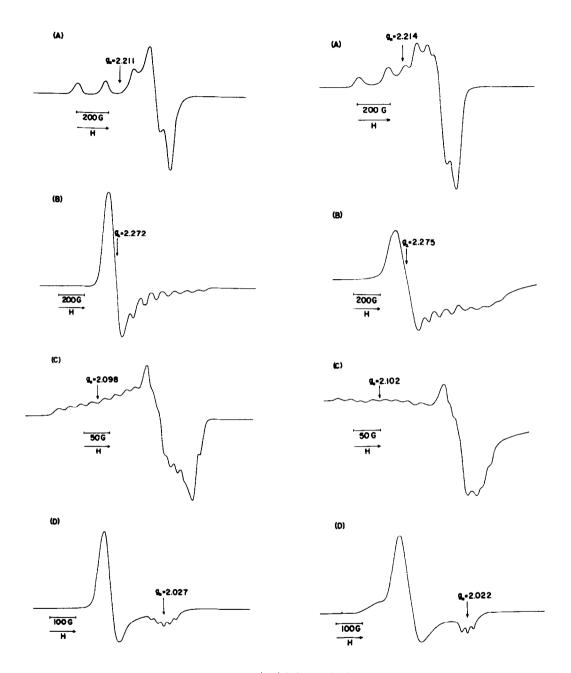


Figure 1 ESR spectra for Cu(II)(A), Co(II)(B), Co(II)-O₂(C), and Ni(III)(D) complexes of BLM-A₂(left) and P-3A(right) at 77 K

reveals lower symmetry of the Cu(II) site(see Figure 1A). Recent X-ray crystallographic result for the 1:1 P-3A-Cu(II) complex indicated a distorted square-pyramidal structure

	Cu(II)			Co(II)			Co(II)-O ₂ g g ₁ A Co,G			Ni(III)		
	g _{ti}	$\mathbf{g}_{\underline{I}}$	A _{II} ^{Cu} ,G	g _{ij}	$\mathbf{g}_{\mathbf{L}}$	A _{II} Co,G	g _{il}	$\mathbf{g}_{\underline{1}}$	A _{ll} Co, G	g _{ij}	$\mathbf{g}_{\underline{\mathbf{I}}}$	A N,G
BLM-A ₂	2.211	2.055	183	2.025	2.272	92.5	2.098	2.007	20.2	2.027	2.169	22.4
P-3A	2.214	2.133 2.078	167	2.027	2.275	93.8	2.102	2.007	22.4	2.022	2.235 2.163	23.5

Table I ESR Parameters for Cu(II), Co(II), Co(II)-0, and Ni(III) Complexes of BLM-A, and P-3A

which involves the secondary amine, pyrimidine, deprotonated peptide, histidine imidazole, and α-amino nitrogen donors. ⁹ The four Cu-N distances of the basal plane range from 1.86 to 2.12 Å, and the fifth axial Cu-N distance is 2.28 Å. Whereas, the sugar and bithiazole portions in BLM-A₂ probably make a contribution to approximately axial symmetry of the 1:1 BLM-A₂-Cu(II) complex.

Of special interest is the fact that P-3A forms low-spin Co(II) and its oxygen adduct complexes similar to BLM-A2. Under the anaerobic condition, the 1:1 P-3A-Co(II) complex also showed the ESR spectrum which is a nearly axial symmetry with the relationship of $g_{\parallel} > g_{\parallel} \simeq 2.0$ and is characteristic of a penta-coordinated square-pyramidal configuration with the unpaired electron in the d₂ orbital(see Figure 1B). 10 By oxygen-bubbling of the P-3A-Co(II) complex, the ESR spectrum drastically changed to that of mono-oxygenated lowspin Co(II) complex(see Figure 1C). The A Co values were estimated to be 15.0 and 16.7 G for the Co(II)-02 complexes of BLM-A2 and P-3A, respectively. A complexes were calculated from the equation of $A_{iso}^{Co} = (A_{\parallel}^{Co} + 2A_{\perp}^{Co})/3$. The A_{\parallel}^{Co} and A_{\perp}^{Co} values were as follows; A_{\parallel}^{Co} =20.2 and A_{\parallel}^{Co} =12.4 G(BLM-A₂ complex) and A_{\parallel}^{Co} =22.4 and A_{\perp}^{Co} =13.8 G(P-3A complex). The effective g values, the relationship of $g_{\parallel} > g_{\parallel} = 2.00$, and the relative small A_{iso} value of these oxygen adduct complexes suggest a considerable delocalization of the unpaired electron from the Co(II) ion. However, it is noted that the oxygenated P-3A-Co(II) complex is more unstable than the BLM-A $_2$ -Co(II)-0 $_2$ complex and undergoes further oxidation to a marked degree. The detailed kinetic and thermodynamic studies of these $Co(II)-0_2$ complex systems are now under way.

Oxidation of the Ni(II) complexes of BLM-A₂ and P-3A in aqueous solution(pH 6.8) yielded paramagnetic products characterized as Ni(III) complexes(see Figure 1D). The ESR results are best described by a tetragonally distorted octahedral geometry with the electron in an orbital which has a large amount of d_Z2 character. The 1:1 BLM-A₂-Ni(III) complex gave the ESR spectrum with $g_{\downarrow} > g_{\parallel}(g_{zz})$ and five-line hyperfine splittings in the g_{\parallel} region. The g_{xx} and g_{yy} values are approximately equal and are not resolved. On the other hand, the ESR spectrum of the 1:1 P-3A-Ni(III) complex showed $g_{xx}, g_{yy} > g_{zz}$ and three-line hyperfine patterns in the g_{zz} region. These results indicate species which have two and one nitrogen nuclei(14N, I=1) bound in the axial position, respectively, for the Ni(III) complexes of BLM-A₂ and P-3A. Complexes of Ni(III) which show strong axial nitrogen coordination have been known to be both kinetically and thermodynamically more stable than those in which the axial donor is water. The stable shows the same of the product of the produc

In conclusion, P-3A is an useful ligand to elucidate the metal-binding sites of BLM in which the secondary amine nitrogen, pyrimidine(N-1) ring nitrogen, deprotonated peptide nitrogen of histidine residue, and histidine imidazole(N-1) nitrogen as planar ligand donors, and the α -amino nitrogen as axial donor, are substantially important. The sugar and bithiazole portions of BLM, however, would contribute to stabilization of the Co(II)-02 adduct complex and axial sixth coordination of the Cu(II) and Ni(III) complexes.

Acknowledgment Gratitude is due to Prof. H. Umezawa for kind encouragement and Dr. T. Takita, Prof. H. Tanaka, and Prof. K. Ishizu for pertinent advice.

References

- 1. Blum, R.H., Carter, S.K., and Agre, K. (1973) Cancer 31, 903-914.
- 2. Umezawa, H., Maeda, K., Takeuchi, T., and Okami, Y. (1966) J. Antibiot. 19A, 200-209.
- 3. Umezawa, H., Suhara, Y., Takita, T., and Maeda, K. (1966) J. Antibiot. 19A, 210-215.
- 4. Dabrowiak, J.C., Greenaway, F.T., Longo, W.E., Husen, M.V., and Crooke, S.T. (1978) Biochim. Biophys. Acta 517, 517-526.
- 5. Sugiura, Y. (1978) J. Antibiot. 31, 1206-1208.
- 6. Chien, M., Grollman, A.P., and Horwitz, S.B. (1977) Biochemistry 16, 3641-3647.

- 7. Kasai, H., Naganawa, H., Takita, T., and Umezawa, H. (1978) J. Antibiot. 31, 1316-1320.
- 8. Bossu, F.P. and Margerum, D.W. (1976) J.Am. Chem. Soc. 98, 4003-4004.
- 9. Iitaka, Y., Nakamura, H., Nakatani, T., Muraoka, Y., Fujii, A., Takita, T., and Umezawa, H. (1978) J.Antibiot. 31, 1070-1072.
- 10. Hoffman, B.M., Diemente, D.L., and Basolo, F. (1970) J. Am. Chem. Soc. 92, 61-65.
- 11. Lappin, A.G., Murray, C.K., and Margerum, D.W. (1978) Inorg. Chem. 17, 1630-1634.